Cell Culture Medium

10% FBS supplemented DMEM

Description

This is the cell culture medium

Ingredients

Add the following solutions to 100ml DMEM

warm up and add 100% FBS 11 ml (10%) - from freezer
penecilin/streptomyosin 1.1 ml (1%) – from freezer
sodium pyruvate 1.1 ml (1%) – from fridge
L-glutamine 1.1 ml (1%) – from freezer.

10% FBS without penecilin/streptomyosin

Description

This is the cell culture medium for transfection. It is the same as the cell culture medium, but without penecilin/streptomyosin.

Ingredients

Add the following solutions to 100 ml DMEM

warm up and add 100% FBS 11ml (10%)
sodium pyruvate 1.1ml (1%)
L-glutamine 1.1ml (1%)

0.5% FBS supplemented DMEM

Description

This is the cell culture medium for starving cells

Ingredients

Add the following solutions to 100ml DMEM

warm up and add 100% FBS 0.5 ml (0.5%)

penecilin/streptomyosin 1.0 ml (1%)

sodium pyruvate 1.0 ml (1%)

L-glutamine 1.0 ml (1%)

0.5% FBS without penecilin/streptomyosin

Description

This is the cell culture medium for starving cells, when the cell condition is not very good.

Ingredients

Add the following solutions to 100 ml DMEM

warm up and add 100% FBS 0.5 ml (0.5%)

sodium pyruvate 1.0ml (1%)

L-glutamine 1.0ml (1%)

CO_2 independent medium with 0.5% FBS

Description

Change cell cultures to this medium right before imaging under microscope

Ingredients

49-50 ml CO_2 independent medium

1 ml L-glutamine (200 mM)

250 ul 100% FBS (0.5%)

0.5X trypsin in PBS (40ml)

warm up and add 2 ml 10X trypsin-EDTA (in freezer) to 38 ml PBS

2% Gelatin (200 ml)

get 200 ml MQ H_2O, weigh 4 g Gelatin (cell culture grade) and add to H_2O, gently shake bottle, put on a tray containing some water, loose the cap of the bottle, autoclave with liquid cycle (30min). After take out, be sure to shake the bottle until gelatin desolve uniformly in liquid, keep in refrigerator for later use.